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(57) Abstract

A nucleic acid fragment comprising a portion of at least 17 contiguous nucleotide bases which portion has a sequence the same as, or homologous to a portion of corresponding length of the sequence of the coding strand as set out in Fig. 1 or the same as, or homologous to a portion of corresponding length of the sequence complementary to the sequence of the coding strand set out in Fig. 1.

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MUCIN NUCLEOTIDES

The present invention relates to nucleotide fragments, polypeptides and antibodies and their use in medical treatment and diagnosis.

In International Patent Application no. WO-A-88/05054 there is disclosed a tandem repeat sequence contained in the human polymorphic epithelial mucin (HPEM) gene and nucleotide probes, polypeptides, antibodies and antibody-producing cells which are useful in the diagnosis and treatment of adenocarcinomas such as breast cancer.

The present inventors have now elucidated the nucleotide base sequence of the gene in the region 5' of the tandem repeat sequence (unless the context implies otherwise, directions such as "5" or "3", "upstream" or "downstream" used herein refer to the non-template strand of the genomic DNA or fragments thereof). The complete sequence of the 1763 nucleotide bases of the non-template strand upstream of and including the first Smal restriction site in the tandem repeat is set out in Fig. The sequence of 1575 nucleotide bases of the nontemplate strand upstream of and including the first Small restriction site in the tandem repeat as set out in Fig. 3 has been extended and some parts have been corrected in the light of repeat experiments. The template strand has a complementary sequence and it is this strand which is transcribed into RNA during expression of the gene

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product.

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In addition to conventional transcriptional and translational start sites and intron splicing sites, this sequence contains a number of features which may be important in the diagnosis and therapy of cancers and in expression of proteins from recombinant vectors. These features will be described below. The amino acid sequence corresponding to the translated portions of this nucleotide sequence gives rise to peptides and thence to antibodies and antibody-producing cells which may also be useful in such diagnosis and treatment.

In one aspect the present invention provides a nucleic acid fragment comprising a portion of at least 17 contiguous nucleotide bases which portion has a sequence the same as, or homologous to a portion of corresponding length of the sequence of the coding strand as set out in Fig. 1 or the same as, or homologous to a portion of corresponding length of the sequence complementary to the sequence of the coding strand set out in Fig. 1.

As used herein the term "fragment" is intended to include restriction endonuclease-generated nucleic acid molecules and synthetic oligonucleotides.

The nucleic acid fragments of the invention may be single-stranded or double-stranded and they may be RNA or DNA fragments. Single stranded fragments may be "plus" or coding strands having the sequence of Fig. 1 or a part

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thereof or a sequence homologous thereto. Alternatively the single stranded fragments may be "minus" or non-coding strands having a sequence complementary to the sequence of Fig. 1 or a part thereof or a sequence homologous thereto. Double stranded fragments contain a complementary pair of strands, (ie. one plus strand and one minus strand).

RNA fragments according to the invention will, of course, contain uridylic acid ("U") residues in place of the deoxythymidylic acid residues ("T") of the coding (non-template) strand set out in Fig. 1 or, if complementary to the sequence of the coding strand, they will contain U residues in positions complementary to the adenylic acid ("A") residues in the coding strand set out in Fig. 1.

15 Preferably the nucleic acid fragments of the invention are double-stranded DNA fragments.

Single-stranded nucleic acid fragments of the invention are at least 17 nucleotide bases in length.

Double-stranded nucleic acid fragments of the invention are at least 17 nucleotide base pairs in length.

Preferably the fragments are at least 20 bases or base pairs in length, more preferably at least 25 bases or base pairs and yet more preferably at least 50 bases or base pairs in length.

Statistically it is almost certain that a 17 nucleotide base sequence will be unique so that any

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nucleic acid fragment having a contiguous portion of 17 nucleotides of a sequence identical to a portion of corresponding length of the coding strand as set out in Fig. 1, or the same as the non-coding strand complementary to the sequence of Fig. 1, will be new. Fragments according to the invention which are only 17 nucleotides or nucleotide bases in length have a sequence the same as, or complementary to, that set out in Fig.1. Longer fragments of the invention may have a sequence which is homologous to a corresponding portion of the sequence for the coding strand as set out in Fig. 1 or to the complementary non-coding strand.

Preferably nucleic acid fragments according to the invention have at least 75% sequence homology with a corresponding portion of the sequence of Fig. 1 or the complementary non-coding strand, for instance 80 or 85%, more preferably 90 or even 95% homology. Differences may arise through deletions, insertions or substitutions. In addition to containing a portion homologous to or the same as the sequence of the coding strand in Fig. 1 or complementary non-coding strand, the nucleic acid fragments of the invention may include sequences completely unrelated to that in Fig. 1.

Particular features of interest within the coding strand in Fig. 1 are set out in Tables 1 to 3 below:

TABLE 1: Signal Sequences

LOCACION~	Sequence in PEM	Significance
1-2	CG	transcriptional start site
73-75	ATG	translational start signal
131-132	GT	start of first intron
631-632	AG	end of first intron
and }	TTCCTGCTGCTGCT- CCTCACAGTGCTTA- CAGTTGTT	Signal sequence, interrupt by first intron (first intindicated by"").
955-960	CCCGGG	Smal site at start of tand repeat
	to Tables 1 and 2 onsensus sequence	•
		•
		s: R is A or G
		s:RisA or G NisA, C, G or T

* Locations are of the 5' base of the indicated PEM sequence numbered as in Fig. 1.

TABLE 2: Regulatory elements within the 5' flanking sequence

Regulatory element	Consensus Sequence+	Sequence in PEM	Location*
SP1	GGGCGG	GGGCGG	
		GGGCGG	-727
		GGGCGG	-397
		GGGCGGCGGGCGGG	-94
	•	GGGCGGGCGGGG	-54
SV40 enhancer element			
a	ATGTGTGT	CTGTGGGT	-562
ъ	GCATGCAT	GCCTGCCT	+25
c	GTGGATAG	GTGGAGAG	- 702
			-102
AP-1	<u>C</u> TGACT <u>C</u> A	GTGACCAC	- 739
	G A	CTGCTTCA	-418
		GTGCCTAG	-61
		CTGCCTGA	+27
			747
AP-2	CC <u>C</u> C <u>A</u> GGC	ACCCAGGC	- 597
	GG	CACCGGGC	+77
			477
IF1/CTF	TTGGCTNNNAGCCAA	TTGGCTTTCTCCAA	-618
33	_		010
Slucocorticoid regulatory			
Core sequence	TGTTCT	TGTTCT	+38
		TGTTCC	- 321
Congonava			722
Consensus sequence	GGTACANNNTGTTCT	GCCTGAATCTGTTCT	+29
		AGCTGGCTTTGTTCC	-330
CACCC factor			
SACCE TACTOR	CACCC	CACCC	+54
		CACCC	+84
Progesterone receptor	3.000.000.000.000		
consensus sequence	ATTCCTCTGT	ACTCCTCTCC	-802
seductice		ACTCCTCCTT	- 626
		ATTTCTCGGC	-432
Estrogen consensus			
sequence	COMOLYMPIA		
·	GGTCANNNTGACC	GCTCCCGGTGACC	-746
RNA Polymerase III			
Box A	Transaction and a second		
2011 11	RRYNNARYXGG	GACCTAGCTGG	- 335
Box B		AGTGGAGTGGG	-388
DON D	GWTCRANNC	GTTCCAGAC	-260
Enhancer sequences:			
Interferon-B seq			
meerreron-p sed	GGAAATTCCTCTG	GGAAATTTCTTCC	-642
CMV enhancer			042
ATT CILIQUEEL	GGAAAGTCCCGTT	GGAAAGTCCGGCT	

The sequence in Fig. 1 also includes two sites occurring in the promoter region and in the first intron having 70 to 80% homology with the mammary consensus, sequence (Rosen, J.M. in "The Mammary Gland, Development, Regulation and Function", Ed. Nevill, M.C. and Daniel, C.W. Plenum Press, pp 301-322). These sites are set out in Table 3 below:

TABLE 3

-289 to -274 AGGCTAAAACT	
* ** * +230 to +245 GTAAGAATTGO	
	
Consensus RGAAGRAAANI	GGACA

Preferred fragments according to the present

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invention include the transcriptional and translational start signals, "TATAA" box and at least one of the regulatory elements (transcription factor binding sites) set out in Table 2 above. More preferably these fragments contain 2 or more, for instance 3, 4 or 5 of the regulatory elements in addition to the TATAA box or even all of the regulatory elements set out in Table 2. Those fragments containing more than one of the regulatory elements of Table 2 preferably also preserve the relative spacings of those sites from one another and from the TATAA box and transcriptional and translational start signals.

Other preferred fragments of the invention contain at least one of the regions homologous to the mammary consensus sequences as set out in Table 3. Preferably these fragments contain both of the regions having homology with the mammary consensus sequences as set out in Table 3. Those fragments containing both regions having homology with the mammary consensus sequence preferably also preserve the relative spacing of those regions, as found in Fig. 1, from one another and from the TATAA box and transcriptional and translational start signals.

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Yet further preferred fragments according to the invention comprise the TATAA box, the transcriptional and translational start signals, at least one and preferably two or more of the regulatory elements as set out in Table 2 and at least one and preferably both of the regions having

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homology with the mammary consensus sequence as set out in Table 3. Yet more preferably these fragments also preserve the relative spacing of the features from Tables 1, 2 and 3. Particularly preferred fragments according to the invention comprise the sequence upstream of the TATAA box as set out in Fig. 1 together with, and downstream thereof, transcriptional and translational start signals and a polypeptide coding sequence in correct reading frame register with the promoter sequences and the TATAA box, transcriptional and translational start signals. The coding sequence may encode a part or parts of the polypeptide encoded by the mucin gene, for instance a part or parts thereof other than the tandem repeat sequence, or polypeptides unrelated to that encoded by the mucin gene.

Other particularly preferred fragments according to the present invention comprise promoter sequences, a TATAA box, transcriptional and translational start signals and, downstream thereof and in correct reading frame register therewith a coding sequence corresponding to a portion of the mucin gene, for instance corresponding to the first exon (corresponding to bases (1 to 130 of Fig.1.) or a part thereof and/or the second exon (corresponding to bases 633 onwards in Fig.1.) or a part thereof, for instance a part thereof other than the tandem repeat sequence as set out in WO-A-88/05054.

In an especially preferred aspect the fragments

contain (i) the first 26 bases (bases 1 to 26 of Fig. 1) or

(ii) the whole of the first exon (bases 1 to 130 of Fig.1.)

and/or (iii) the splicing/ligating sites for the first intron

set out in Table 1 and a non-coding sequence between these

5 sites. The non-coding sequence may be the same as or

different to the sequence of the first intron as shown in

Fig. 1. Preferably it is the same.

Other preferred fragments of the invention comprise at least a portion of the first intron (bases 231 to 632 of Fig. 1). Further preferred fragments of the invention comprise at least a portion of the 5'-flanking sequence upstream of base -423 of Fig. 1.

Other preferred fragments of the invention comprise a portion of the sequence of Fig. 1 corresponding to a portion of the sequence of Fig. 3.

Further preferred fragments of the invention comprise a combination of any two or more of the foregoing preferred features.

Fragments according to the present invention

containing functional coding sequences for a least a part of
the first or second exons set out in Fig. 1 are useful in the
production of polypeptides corresponding to a part or all of
the mucin gene product. Such polypeptides are, in turn
useful as immunogenic agents for instance in active

immunisation against Human Polymorphic Epithethial Mucin

(HPEM) for the prophylactic or therapeutic treatment of

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cancers or raising antibodies for use in passive immunisation and diagnosis of cancers. For use in such methods the fragment, which codes for a polypeptide chain substantially identical to a portion of the mucin core protein, may be 5 extended at either or both the 5' and 3' ends with further coding or non-coding nucleic acid sequence including regulatory and promoter sequences, marker sequences, and splicing or ligating sites. Coding sequences may code for other portions of the mucin core protein chain (for instance, 10 other than the tandem repeat) or for other polypeptide The fragment according to the invention, together with any necessary or desirable flanking sequences is inserted, in an appropriate open reading frame register, into a suitable vector such as a plasmid, or cosmid or a viral 15 genome (for instance vaccinia virus genome) and is then expressed as a polypeptide product by conventional techniques. In one aspect the polypeptide product may be produced by culturing appropriate cells transformed with a vector, harvested and used as an immunogen to induce active 20 immunity against the mucin core protein [Tartaglia et al., <u>Tibtech</u>, <u>6</u>, 43: (1988)].

Fragments according to the present invention incorporating regulatory elements of Table 2 and/or mammary consensus sequences of Table 3 may be used in securing tissue-specific expression of functional coding sequences in appropriate reading frame register downstream of the

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regulatory elements and/or associated with the mammary consensus sequences. Such fragments may therefore be used to express parts or the whole of the mucin gene or any other coding sequence in cells of epithelial origin. Applications of this are in therapy and immunisation where such fragments and associated coding sequences are administered to patients such that the coding sequence will be expressed in epithelial tissues leading to a therapeutic effect or an immune reaction by the patient against the polypeptides.

The fragments may be presented as inserts in a vector such as viral genomic nucleic acid and introduced into the patients by inoculation of the vector for instance as a modified virus. The vector then directs expression of the polypeptide <u>in vivo</u> and this in turn serves as a therapeutic agent or as an immunogen to induce active immunity against the polypeptide. This strategy may be adopted, for instance, to secure expression of polypeptides encoded by the HPEM gene for treatment or prophylaxis of adenocarcinomas such as breast cancer or to secure tissue specific expression of other peptides under control of the regulatory sequences of Table 1, for instance by administration of a modified vaccinia virus containing the fragment and coding sequences in its genomic DNA. RNA fragments of the invention may similarly be used by administration via a retroviral vector. Selection of tissue specific virus vectors to carry the fragments of the

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invention and coding sequences will further restrict expression of the polypeptide to desired target tissues.

Fragments of the invention may also be used to control expression of oncogenic proteins in experimental transgenic animals. Thus, for instance, a transgenic mouse having an oncogene such as ras, erbB-2 or int 2 expressed under control of the present tissue specific fragments may develop breast tumours and be useful in testing diagnostic agents such as tumour localisation and imaging agents and in testing therapeutic agents such as immunotoxins. Nucleic acid fragments according to the invention are also useful as hybridisation probes for detecting the presence of DNA or RNA of corresponding sequence in a sample. For use as probes fragments are preferably labelled with a detectable label such as a radionuclide, enzyme label, fluorescent label or other conventional directly or indirectly detectable labels. For some applications, the probes may be bound to a solid support. Labelling of the probes may be achieved by conventional methods such as set out in Matthews et al., Anal. Biochem.

<u>169:</u> 1-25 (1988).

In further aspects, the present invention provides cloning vectors and expression vectors containing fragments according to the present invention. The vectors may be, for instance, plasmids, cosmids or viral genomic DNA. The present invention further provides host cells containing

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such cloning and expression vectors, for instance epithelial cells transformed with functional expression vectors containing expressible fragments according to the invention.

The invention further provides nucleic acid fragments, which encode polypeptides as defined below. Such fragments may be fragments as hereinbefore defined. However, in view of the redundancy of the genetic code, nucleic acid sequences which differ slightly or substantially from the sequence of Fig. 2 may nevertheless encode the same polypeptide.

The nucleic acid fragments of the invention may be produced <u>de novo</u> by conventional nucleic acid synthesis techniques or obtained from human epithelial cells by conventional methods, Huynh <u>et al.</u>, "DNA Cloning: A Practical Approach" Glover, D.M. (Ed) IRL, Oxford, Vol 1, pp49-78 (1985).

The invention therefore also provides probes, vectors and transformed cells comprising nucleic acid fragments as hereinbefore defined for use in methods of treatment of the human or animal body by surgery or therapy and in diagnostic methods practiced on the human or animal body and for use in the preparation of medicaments for use in such methods. The invention also provides methods for treatment of the human or animal body by surgery or therapy and diagnostic methods practiced in vivo as well as ex vivo and in vitro which comprise administering such fragments, probes, vectors or

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transformed cells in effective non-toxic amount to a human or other mammal in need thereof.

Processes for producing fragments according to the invention and probes, vectors and transformed cells

5 containing them and processes for expressing polypeptides encoded by, or under the regulatory control of, fragments of the invention also form aspects of the invention.

The invention further provides a polypeptide comprising a sequence of at least 5 amino acid residues encoded by the coding portion of the DNA sequence as indicated in Fig. 2. Polypeptides according to the invention preferably have a sequence of at least 10 residues, for instance at least 15, more preferably 20 or more residues and most preferably all the residues shown in Fig. 2.

The polypeptide may additionally comprise N-terminal and/or C-terminal sequences not encoded by the DNA sequence indicated by Fig. 2.

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Polypeptides of the invention containing more than 5 amino acid residues encoded by the DNA sequence in Fig. 2 may include minor variations by way of substitution, deletion or insertion of individual amino acid residues. Preferably such polypeptides differ at not more than 20% preferably not more than 10% and most preferably not more than 5% of residues in a contiguous portion corresponding to a portion of the sequence in Fig. 2.

The invention further provides polypeptides as

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defined above modified by addition of a linkage sugar such as N-acetyl galactosamine on serine and/or threonine residues and polypeptides modified by addition of oligosaccharide moieties to N-acetyl galactosamine or via other linkage sugars. Optionally modified polypeptides linked to carrier proteins such as keyhole limpet haemocyanin, albumen or thyroglobulin are also within the invention.

Polypeptides according to the invention may be 10 produced de novo by synthetic methods or by expression of the appropriate DNA fragments described above by recombinant DNA techniques and expressed without glycosylation in human or non-human cells. Alternatively they may be obtained by deglycosylating native human mucin glycoprotein (which 15 itself may be produced by isolation from samples of human tissue or body fluids or by expression and full processing in a human cell line) [Burchell et al., Cancer Research, 47: 5467-5482, (1987), Gendler et al., P.N.A.S., 84: 6060-6064, (1987)], and digesting the core protein. The polypeptides 20 of the invention are useful in active immunisation of humans, for raising antibodies in animals for use in passive immunisation, diagnostic tests, tumour localisation and, when used in conjunction with a cytotoxic agent, for tumour therapy.

The invention further provides antibodies against any of the polypeptides described above.

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As used hereafter the term "antibody" is intended to include polyclonal and monoclonal antibodies and fragments of antibodies bearing antigen binding sites such as the F(ab')₂ fragments as well as such antibodies or fragments thereof which have been modified chemically or genetically in order to vary the amino acid residue sequence of one or more polypeptide chains, to change the species specific and/or isotype specific regions and/or to combine polypeptide chains from different sources. Especially in therapeutic applications it may be appropriate to modify the antibody by coupling the Fab, or complementaritydetermining region thereof, to the Fc, or whole framework, region of antibodies derived from the species to be treated (e.g. such that the Fab region of mouse monoclonal antibodies may be administered with a human Fc region to reduce immune response by a human patient) or in order to vary the isotype of the antibody (see EP-A-0 239 400). Such antibodies may be obtained by conventional methods [Williams, Tibtech, 6:36, (1988)] and are useful in diagnostic and therapeutic applications, such as passive immunisation.

The term "antibodies" used herein is further intended to encompass antibody molecules or fragments thereof as defined above produced by recombinant DNA techniques as well as so-called "single domain antibodies" or "dAbs" such as are described by Ward, E.S. et al., Nature, 341:544-546

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(1989) which are produced in recombinant microorganisms, such as Escherichia coli, harboring expressible DNA sequences derived from the DNA encoding the variable domain of an immunoglobulin heavy chain by random mutation introduced, for instance, during polymerise chain reaction amplification of the original DNA. Such dAbs may be produced by screening a library of such randomly mutated DNA sequences and selecting those which enable expression of polypeptides capable of specifically binding the polypeptides of the invention or HPEM core protein.

Antibodies according to the present invention react with HPEM core protein, especially as expressed by colon, lung, ovary and particularly breast carcinomas, but have reduced or no reaction with corresponding fully processed HPEM. In a particular aspect the antibodies react with HPEM core protein but not with fully processed HPEM glycoprotein as produced by the normal lactating human mammary gland.

Antibodies according to the present invention preferably have no significant reaction with the mucin glycoproteins produced by pregnant or lactating mammary epithelial tissues but react with the mucin proteins expressed by mammary epithelial adenocarcinoma cells. These antibodies show a much reduced reaction with benign breast tumours and are therefore useful in diagnosis and localisation of breast cancer as well as in therapeutic methods.

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Further uses of the antibodies include diagnostic tests of assays for detecting and/or assessing the severity of breast, colon, ovary and lung cancers.

The antibodies may be used for other purposes including screening cell cultures for the polypeptide expression product of the human mammary epithelial mucin gene, or fragments thereof, particularly the nascent expression product. In this case the antibodies may conveniently be polyclonal or monoclonal antibodies.

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The invention further provides antibodies linked to therapeutically or diagnostically effective ligands. For therapeutic use of the antibodies the ligands are lethal agents to be delivered to cancerous breast or other tissue in order to incapacitate or kill transformed cells. Lethal agents include toxins, radioisotopes and "direct killing agents" such as components of complement as well as cytotoxic or other drugs.

For diagnostic applications the antibodies may be linked to ligands such as solid supports and detectable labels such as enzyme labels, chromophores, fluorophores and radioisotopes and other directly or indirectly detectable labels. Preferably monoclonal antibodies are used in diagnosis.

Antibodies according to the present invention may be produced by inoculation of suitable animals with a polypeptide as hereinbefore described. Monoclonal

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antibodies are produced by known methods, for instance by the method of Kohler & Milstein [Nature, 256: 495-497 (1975)] by immortalising spleen cells from an animal inoculated with the mucin core protein or a fragment thereof, usually by fusion with an immortal cell line (preferably a myeloma cell line), of the same or a different species as the inoculated animal, followed by the appropriate cloning and screening steps.

Antibody-producing cells obtained from animals inoculated with polypeptides of the invention and immortalised such cells form further aspects of the invention.

The invention further provides polypeptides, antibodies and antibody producing cells, such as hybridomas, as hereinbefore defined for use in methods of surgery, therapy or diagnosis practiced on the human or animal body or for use in the production of medicaments for use in such methods. The invention also provides a method of treatment or diagnosis which comprises administering an effective non-toxic amount of a polypeptide or antibody as hereinbefore described to a human or animal in need thereof.

Processes for producing polypeptides according to the invention whether by expression of nucleic acid fragments of the invention of otherwise, and for producing antibodies or fragments thereof and for producing antibody-producing cells such as immortalised cells, form further aspects of the

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invention.

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The invention further provides a diagnostic test or assay method comprising contacting a sample suspected to contain abnormal human mucin glycoproteins with an antibody as defined above. Such methods include tumour localisation involving administration to the patient of the antibody bearing detectable label or administration of an antibody and, separately, simultaneously or sequentially in either order, administering a labelling entity capable of selectively binding the antibody or fragment thereof. Diagnostic test kits are provided for use in diagnostic tests or assays and comprise antibody and, optionally, suitable labels and other reagents and, especially for use in competitive assays, standard sera.

The invention will now be illustrated with reference to the figures of the accompanying drawings in which:

Fig. 1. shows the deoxynucleotide base sequence of the 1763 bases upstream of and including the first SmaI restriction site in the tandem repeat sequence of WO-A-88/05054 using the conventional symbols A, C, G and T for the bases of the non-template strand. The base sequence is arranged in blocks of ten. Untranscribed sequence is in lower case, transcribed sequence is in upper case. The SP1 regulatory elements (Table 2), TATAA box, transcriptional and translational start sites (Table 1) are underlined.

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Fig. 2. shows the sequence of the non-template strand commencing from the transcriptional start site, (residue 1 in Fig. 1.) and excluding the sequence of the first intron (bases 131 to 632 of the sequence in Fig.1.). Fig.2 also shows the predicted sequence of the polypeptide using the conventional 1 letter symbols for the amino acid residues. Amino acid residues are numbered down the left-hand side and nucleotide bases down the right hand side. The signal sequence is underlined. The sequences end at the first SmaI site in the tandem repeat.

Fig. 3. shows the deoxy nucleotide base sequence of the 1575 bases upstream of and including the first SmaI restriction site in the tandem repeat sequence of WO-A-88/05054 using

15 the conventional symbols A, C, G and T for the bases of the non-template strand. The base sequence is arranged in blocks of ten in non-coding regions. The exon sequences are shown in blocks of three and translated codons are underlined. The start positions of exons 1 and 2, intron 1 and the signal sequence for exon splicing are numbered and labelled. Other features mentioned in Tables 1 and 2 are boxed. The sequence finishes with the first SmaI site of the tandem repeat sequence.

The present invention does not extend to fragments,

polypeptides and antibodies or related materials such as vectors and cells, which are specifically disclosed in WO-A-88/05054 or WO-A-90/05142, nor to the CDNA fragment whose sequence is indicated in Abe, M. et al., in Biochemical and Biophysical Research Communications, 165(2): 644-649 (1989).

The invention will now be illustrated by the following Examples:

EXAMPLE 1

In an attempt to obtain clones with 5' unique 10 sequences, two gt10 libraries were screened with a probe for the tandem repeat. All the clones obtained lacked any non-repetitive sequence at the 5' terminus. Thus, a different strategy was adopted. To obtain 5' sequence we synthesized the cDNA corresponding to the 5' end of breast 15 cancer cell line transcript using anchored-polymerise chain reaction (A-PCR). The A-PCR procedure [Loh, E.Y. et al., Science, 243: 217-220, (1989) was used to synthesize cDNA corresponding to the 5' end of the transcript. For the 5' end clones total RNA (5 μ g) prepared by the guanidinium isothiocyanate method [Chirgwin, J.M. et al., Biochem., 18: 20 5294-5299 (1979)] was used for first strand synthesis using a breast cancer cell line (BT20) transcript with AMV-reverse transcriptase (Life Sciences) in a 40 μ l reaction mixture

[Okayama, H. and Berg, P., Mol. Cell. Biol., 2: 161-170 (1982)] containing 1 μ g of an oligonucleotide primer made to the tandem repeat (5'CCAAGCTTGGAGCCCGGGGCCGGCCTGGTGTCCGG3'). The total RNA was subjected to reverse transcription, and the products were precipitated with spermine. A poly(dG) 5 tail was introduced with terminal deoxy-transferase (500 U/ml, Pharmacia). Amplification was performed with Thermus aquaticus polymerise (Perkin Elmer Cetus) in 100 μ l of the standard buffer supplied. The primers included the tandem repeat primer and for the poly(dG) end, a mixture of the AN 10 the AN primer (5'GCATGCGCGCGGCGGGGGCC3') at a ratio of 1:9. Following an initial denaturation at 94°C for 5 min, the reaction was annealed at 55°C for 2 min, extended at $72^{\rm O}{\rm C}$ for 2.5 min and denatured at 94°C for 1.5 min. 15 Amplification was performed for 30 cycles, and the product was precipitated with ethanol. The DNA was sequentially cut with HindIII and SacII, separated on a 1.2% agarose Gel and the band of approximately 550 bp was purified onto DEAE membrane (Schleicher and Schuell), ligated into pBS-SK+ and 20 transformed into bacteria XL-1 (Stratagene). This plasmid will be referred to as pBS-5'PEM. All restriction enzymes used were obtained from New England Biolabs Inc., oligonucleotide primers and probes were synthesized on an Applied Biosystems 380B DNA synthesizer. 25

Four colonies were selected for sequencing, and the

sequences agreed with each other and with sequence obtained from genomic clones of the region. A Leader sequence of 72 bp preceded the first ATG which was in-frame with the reading frame of the tandem repeat as previously determined (Fig. 1), and the sequence preceding first ATG, CCACCATGA, agrees with the Kozak consensus sequence (Kozak, M., Nucl. Acids. Res., 12: 857-872 (1984).

The primer extension technique was use to map precisely the position of the capsite. A 21 bp 10 oligonucleotide primer (5'AGACTGGGTGCCCGGTGTCAT3') corresponding to nucleotides 73 to 93 ending at the A of ATG (Fig. 1) was end-labelled with $[\forall -3^2P]$ ATP (> 5000 Ci/mmol, Amersham International plc) using T4 poly-nucleotide kinase (Pharmacia) and precipitated three times with equal volumes of 4 M ammonium acetate to remove free [%-32P]ATP from the 15 kinased oligonucleotide. Labelled primer (1 x 10^5 dpm at 1 x 10 7 dpm/pmole) was annealed to 40 μ g of total BT 20 RNA in 120 mM sodium chloride at 95°C for 5 min, held at 65°C for 1 h and cooled to room temperature. The annealed primer was extended using 18 units of reverse transcriptase in 50mM 20 Tris pH 8.3 at 45°C, 6 mM magnesium acetate, 10 mM dithiothreitol, 1.8 mM dNTPs in a total volume of 50 μ l at 45°C for 1h. The reaction was stopped by the addition of 50 $m\underline{M}$ EDTA and the RNA digested by treatment with RNase-A at 25 $400\mu g/ml$ for 15 min at 37°C. The samples were than phenol:chloroform extracted prior to ethanol precipitation

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and electrophoresed on a standard 6% sequencing gel yielding two bands which mapped to two C's, 72 and 71 bases upstream of the ATG. The sequencing ladder was single-stranded control DNA (M13mp18) from the Sequenase kit (US Biochemical Corp.).

The most prominent product was 72 bp, equal to the number of base pairs from the 5' end of the oligonucleotide primer to the 5' end of the PCR-derived clone, thus confirming that the cDNA represents the entire length of its corresponding cellular mRNA 5' to the tandem repeat. The presence of a second band may be due to interference with reverse transcriptase by methylation of the C at base 71, since it forms a CpG dinucleotide. Under identical conditions, no primer extension product was seem using RNA from Daudi cells which do not express the PEM mucin.

Cloning

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A plasmid library, grown in DH1\(\alpha\)cells (RecA-), was used instead of a lambda library, because of the possibility of recombination occurring when lambda is grown in RecA+cells. This recombination might have been expected, since a part of the tandem repeat sequence (GCTGGGGG) is closely related to the chi sequence (GCTGGTGG) of lambda phage which has been implicated as a hotspot for RecA-mediated recombination in E.coli.

Nucleotide sequence of cDNA clones

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Fig 1. shows the DNA sequence from the 5'

A-PCR-derived clone, including the consensus sequence of the tandem repeat. Sequences were determined in both

directions. The region of conserved tandem repeats was not sequenced in full, although a cDNA tandem repeat clone obtained previously had been circularised, sonicated and about 40 clones sequences [(Gendler et al., J. Biol. Chem., 263:12820-12823 (1988)].

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<u>Predicted amino acid sequence and composition of the PEM core protein.</u>

The core protein amino acid composition is dominated by the amino acid composition of the tandem repeat. Serine, threonine, proline, alanine and glycine account for about 60% of the amino acids.

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The deduced sequence of the PEM core protein consists of distinct regions including (1) the N-terminal region containing a hydrophobic signal sequence and degenerate tandem repeats and (2) the tandem repeat region itself. At the N-terminus a putative signal peptide of 13 amino acids follows the first 7 amino acids. However, the actual site of cleavage has not been determined as attempts to obtain N-terminal sequence of the core protein were hindered by a blocked amino terminus. Following the signal sequence and preceding the first SmaI site (which is used to define the beginning of the tandem repeat region) are 107 amino acids. Greater than 50% of these amino acids comprise degenerate tandem repeats. Since the number of tandem repeats per molecule is large (greater than 21 for the smallest allele we have observed), this domain forms the major part of the core protein, and results in a highly repetitive structure which is extremely immunogenic

25 [Gendler, S. et al., loc. cit]. The sequence of the 20

amino acid tandem repeat unit corresponds to what might be expected for a protein which is extensively 0-glycosylated. Five serines and threonines, four of which are in doublets, are found in the repeat and these potential glycosylation sites are separated by regions rich in prolines (See Fig. 2).

CLAIMS

- A nucleic acid fragment comprising a portion of at least 17 contiguous nucleotide bases which portion has a sequence the same as, or homologous to a portion of
 corresponding length of the sequence of the coding strand as set out in Fig. 1 or the same as, or homologous to a portion of corresponding length of the sequence complementary to the sequence of the coding strand set out in Fig. 1.
- 2. A fragment according to claim 1 comprising any one or 10 more of the following:
 - (a) a signal sequence

 TTCCTGCTGCTGCTCCTCACAGTGCTTACAGXTTGTT

 wherein X is an optionally present intron
 - (b) a mammary consensus sequence AGGCTAAAACTAGACC
- 15 (c) a mammary consensus sequence GTAAGAATTGCAGACA
 - (d) a homologue of a sequence (a), (b) or (c) and
 - (e) a sequence complementary to a sequence (a), (b), (c)
 or (d).
- A hybridisation probe comprising a fragment according
 to claim 1 or claim 2 bearing a detectable label or linked to a solid support.
 - A cloning or expression vector comprising a fragment

according to claim 1 or claim 2.

- 5. A transformed cell comprising a cloning or expression vector according to claim 4.
- A polypeptide comprising a sequence of at least 5
 contiguous acid residues encoded by the coding portion of the
 DNA sequence as indicated in Fig. 2.
 - 7. An antibody against a polypeptide according to claim 6.
- 8. An antibody according to claim 7 bearing a detectable 10 label or linked to a solid support.
 - 9. An antibody-producing cell capable of secreting an antibody according to claim 7.
- 10. A diagnostic kit comprising a fragment according to claim 1 or claim 2 or a probe according to claim 3 or a
 15 polypeptide according to claim 6 or an antibody according to claim 7 or claim 8.
 - 11. A fragment according to claim 1 or claim 2 or a probe according to claim 3 or a vector according to claim 4 or a cell according to claim 5 or claim 9 or a polypeptide

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according to claim 6 or an antibody according to claim 7 or claim 8 for use in a method of treatment or diagnosis practised on the human or animal body.

- 12. Use of a fragment according to claim 1 or claim 2 or a probe according to claim 3 or a vector according to claim 4 or a cell according to claim 5 or claim 9 or a polypeptide according to claim 6 or an antibody according to claim 7 or claim 8 in the preparation of a medicament for use in a method of treatment or diagnosis practised on the human or animal body.
- 13. A method of treatment or diagnosis comprising administering to a cancer patient in need thereof or suspected to have a cancer an effective non-toxic amount of a fragment according to claim 1 or claim 2 or a probe according to claim 3 or a vector according to claim 4 or a cell according to claim 5 or claim 9 or a polypeptide according to claim 6 or an antibody according to claim 7 or claim 8.
- 14. A method of diagnosis comprising contacting a sample from a patient with a fragment according to claim 1 or claim 20 2 or a probe according to claim 3 or a vector according to claim 4 or a cell according to claim 5 or claim 9 or a polypeptide according to claim 6 or an antibody according to claim 7 or claim 8.

ggggaggag cccaaaacta gcacctagtc cactcattat ccagccctct tatttctcgg ccgctctgct tcagtggacc tactcctctc cgcccggtcc gagcggccc tcagcttgcg cggcccagcc ccgcaaggct cccggtgacc actagagggc <u>gg</u>gaggagct cctggccagt ggtggagagt ggcaaggaag gaccctaggg ttcatcggay cccaggttta ctccttaag tggaaattte tteceecaet eeteettgge ttteteeaag gagggaaeee aggetgetgg aaagteegge tggggggggg actgtgggtt cagggggagaa cgggggtgtgg aacgggacag ggagcggtta gaagggtggg gctattccgg gaagtggtgg cgggga<u>gggc gg</u>ggaagtgg agtgggagac ctaggggtgg gcttcccgac cttgctgtac aggacctcga cctagctggc tttgttcccc atccccacgt tagttgttgc cctgaggcta aaactagagc ccaggggccc caagttccag actgccctc cccctcccc cggagccagg gaglggttgg tgaaaggggg aggccagctg gagaacaaac gggtagtcag ggggttgage gattagagcc cttgtaccct acccaggaat ggttggggag gaggaggaag aggtaggagg taggggaggg ggcggggttt tgicaccigi caccigcicg cigigcciag <u>ggcgggcggg cggg</u>gagigg ggggaccgg<u>i ataa</u>agcggi aggcgccigi gccCGCTCCA CCTCTCAAGC AGCCAGCGCC TGCCTGAATC TGTTCTGCCC CCTCCCCACC CATTTCACCA CCACCATGAC -103

Fig.

Fig.1 Cont'd

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157	TCTTCCTGCT GCTGCTCCTC ACAGTGCTTA CAGGtgaggg gcacgaggtg gggagtgggc		cgiggiciti cigigggiti igciccigg cagaiggcac caigaagita aggiaagaai		ccaaccctga		agaggaagg cttaaqaqq		agccaaggcc tatgggcaga gagaaggagg ctgctgcagg gaaggaggct tccaacccag	+557	ttcttgctct		aacagTTGTT)	TGCCCAGCTC		ACCACTCAGG		CACCTCGGTC	+957	AGCCAGCCCC
	gcacgaggtg		catgaagtta	+307	agagttgccc				gaaggaggct		ttatattttc		ctaaacccgc	+707	AGAAGTTCAG		AGGCTCCTCC		GACAGGATGT		CCGGACAACA
	CAGgtgaggg	7	cagatggcac		tctgtgccag aaggaggag aggctaagga caggctgaga		gagagtaggg	+457	ctgctgcagg		tececagtee tectggtatt atttetetgg tggecagage ttatatttte ttettgetet		ccaaccctat gactttaact tcttacagct accacagccc ctaaacccgc aacaqTTGTT	٠	CTCTACCCCA GGTGGAGAAA AGGAGACTTC GGCTACCCAG AGAAGTTCAG TGCCCAGCTC		GTATGACCAG CAGCGTACTC TCCAGCCACA GCCCCGGTTC AGGCTCCTCC ACCACTCAGG	+857	CCGCCCACGG AACCAGCTIC AGGITCAGCI GCCACCIGGG GACAGGAIGT CACCICGGIC		GGGCTCCACC ACCCGGCCAG CCCACGATGT CACCTCAGCC CCGGACAACA AGCCAGCCCC
	ACAGTGCTTA	+207	tgctccctgg		aggctaagga		ctagggggaa		gagaaggagg		atttctctgg	+607	tcttacagct		AGGAGACTTC		TCCAGCCACA		AGGTTCAGCT		CCCACGATGT
	GCTGCTCCTC		ctgtgggttt		ааддаддад	+357	caaatgtcct gtagagaagt		tatgggcaga		tcctggtatt		gactttaact		GGTGGAGAAA	+757	CAGCGTACTC		AACCAGCTTC		ACCCCCCCAG
107	TCTTCCTGCT	: : :	cgiggictit		tctgtgccag		caaatgtcct		agccaaggcc	+207	tccccagtcc		ccaaccctat		CTCTACCCCA		GTATGACCAG		CCGGCCACGG	+907	GGGCTCCACC.
	ACCGGGCACC CAGTCTCCTT	4	ugcccugctc aggrggtctt ±267	167	lgcagacaga ggctgccctg		gagtgggtac cagggggcaag		aggggccatg		gggttactga ggctgcccac		tattttcct tcataaagac	+657	ACAGGITCIG GICAIGCAAG		TACTGAGAAG AATGCTGTGA		GACAGGATGT CACTCTGGCC		GGCCAGCCCT
	ACCGGGCACC	+ 00000+	1106100061	7	tgcagacaga		gagtgggtac	+407	gaagaaatgc		gggttactga		tatttttcct		ACAGGTTCTG		TACTGAGAAG	+807	GACAGGATGT		CCAGTCACCA GGCCAGCCCT

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36	96	156	216	3/6 319	336	396	456	
CCG CTC CAC CTC TCA AGA GCC AGC GCC TGC CTG AAT 36	TTC TGC CCC CTC CCC ACC CAT TTC ACC ACC ACC ATG ACA CCG GGC ACC CAG TCT CCT 96 T P G T Q S P	SIGNAL SEQUENCE TTC CTG CTG CTC CTC ACA GTG CTT ACA GGT TCT GGT CAT GCA AGC 156 F L L L L T V L T V V T G S G H A S	ACC CCA GGT GGA GAG ACT TCG GCT ACC CAG AGA AGT TCA GTG CCC AGC TCT 216 T P G G E K E T S A T Q R S S V P S S	GAG AAG AAT GCT GTG AGT ATG ACC AGC GAT GCT CAC AGC CAC AGC CCC GGT TCA 276 E K N A V S M T S S V L S S H S P G S	TCC TCC ACC ACT CAG GAT GTC ACT CTG GCC CCG GCC ACG GAA CCA GCT TCA 336 S S T T Q G Q D V T L A P A T E P A S	TCA GCT GCC ACC TGG GGA CAG GAT GTC ACC TGG GTC CCA GTC AGG CCA GCC CTG 396 S A A T W G O D V T S V P V T R P A L	TCC ACC ACC CCA GCC CAG GAT GTC ACC TCA GCC CCG GAC AAG CCA GCC CCG 456 S T T P P A H D V T S A P D N K P A P	Fig. 2
	CTG T	TTC T	TCT A 28 S	ACT T	9 099	667	96C . 6	000 000 000
			77	8 7	68	88	108	

SUBSTITUTE SHEET

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.468 3AGC	-398 ACCC	-328 FCGA	.258 3000	.188 3AGA	.118 \GGA	.48 .66
GGGAGGG	CAGTGGA	AGGACCT	GCAGGGG	CAGCTGG	GGGAGGA	:48 CCTAGGGCGG
AAGTGGTGGG	0601016011	CTTGCTGTAC	AAACTAGACC	AAGGGGAGGC	AGGAATGGTT	6010601616
GCTATTCCGG	ATTTCTCGGC	GCTTCCCGAC	CCTGAGGCTA	TGGTTGGTGA	TACCCTACCC	CCTGTCACCT
GAAGGGTGGG	CAACCGTCTT	CTAGGGGTGG	TAGTTĢTTGC	AGCCAGGGAG	AGAGCCCTTG	GGTTTTGTCA
GGAGCGGTTA	ACTCATTATC	AGTGGGAGAC	ATCCCCACGT	CCCTCCCCCGGG	TTGAGCAGTT	GAGGGGGCGG
GAACGGAACG	CACCTAGTCC	GGGGAAGTG	TTTGTTCCCC	ACTGCCCTCC	AGTCAGGGGG	GGAGGTAGGG
-537 ACTGCGTGTG	-467 CCAAAACTAG	.388 GGGGAGGGGC	·327 CCTAGCTGGC	·257 CAAGTTCCAG	·187 ACAAACGGGT	.117 GGAAGAGGTA
	CGTGTG GAACGGAACG GGAGCGGTTA GAAGGGTGGG GCTATTCCGG AAGTGGTGG GGGAGG	SAACG GGAGCGGTTA GAAGGGTGGG GCTATTCC <mark>GG AAGTG</mark> GTGGG GGGAGG	SAACG GGAGCGGTTA GAAGGGTGGG GCTATTCCGG AAGTGGTGG GGGAGGGAGCC AGTCCATTATC CAACCGTCTT ATTTCTCGGC CGCTCTGCTT CAGTGGAGC CTGGGGGGGGG GCTTCCCGAC CTTGCTGTAC AGGACC	SAACG GGAGCGGTTA GAAGGGTGGG GCTATTCCGG AAGTGGTGGG GGGAGGGAGTCC ACTCATTATC CAACCGTCTT ATTTCTCGGC CGCTCTGCTT CAGTGGAGTCG AGTGGCAGAGTGGAGGGGGGGGGG	SAACG GGAGCGGTTA GAAGGGTGGG GCTATTCCGGG AGTCC ACTCATTATC CAACCGTCTT ATTTCTCGGC AGTGB AGTGGAGAC CTAGGGGTGG GCTTCCCGAC TCCCC ATCCCCACGT TAGTTGTTGC CCTGAGGCTA SCTCC CCCTCCGCGG AGCCAGGGAG TGGTTGGTGA	SAACG GGAGCGGTTA GAAGGGTGGG GCTATTCCGGG AGTCC ACTCATTATC CAACCGTCTT ATTTCTCGGC AGTGB AGTGGAGAC CTAGGGGTGG GCTTCCCGAC TCCCC ATCCCCACGT TAGTTGTTGC CCTGAGGCTA SCTCC CCCTCCCGCG AGCCCAGGGAG TGGTTGGTGA SGGGG TTGAGCAGTT AGAGCCCTTG TACCCTACCC

Fig.

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1 CCG CTC CAC CTC TCA ACA GCC	73 CTG TTC TGC CCC CTC CCC ACC CAT TTC ACC ACC ACC ATG ACA CCG GGC ACC CAG	131 INTRON 1 GT GAGGGCAC	c itteteteee cittiectee	C CCTGTCTGTG CCAGAAGGAG	G GTACCAGGGG CAAGCAAATG	G AGGGGAAGAA ATGCAGGGGC	A GGCTTCCAAC CCAGGGGTTA	
1 CCTGTGC	TTC ACC ACC	9	1061166110	CAGAGGCTGC	CTGAGAGTGG	AAGGCTTAAG	CAGGGAAGGA	(1)
GCGGTAGGCG	CCC ACC CAT	A GTG CTT ACA	AGGTTGGTCT	GTTAAGGTAA GAATATCAGA	GCCCCCAACC	AGGGAGAGGG	GAGGCTGCTG	Fig. 3 (ont'd(1)
CCGGTATAAA GCGGTAGGCG	TC TGC CCC CTC	VAL SEQUENCE CTG CTG CTC ACA GTG CTT ACA G	6000116011	GTTAAGGTAA	GAGAAGAGTT	GGAAGAGAGT	CAGAGAGAG GAGGCTGCTG	Fig.
AGTGGGGGGA		SIGNAL SEQUENCE	GAGGTTGGGG AGTTGGGCTT	GCACCATGAA	AGGACAGGCT	AAGTCTAGGG	GGCCTATGGG	
60000000	AGC GCC TGC CTG AAT	97 S1GN 1CI CCI 11C 11C CTG	6466116666	CTGGCAGATG	GGAGGGCTA	TCCTGTAGAG	CATGAGCCAA	

CTGGTGGCCA GAGCTTATAT TTTCTTCTTG GCCCCTAAAC OTCTIATITI TCCTTCATAA AGACCCAACC CTATGACTTT AACTTCTTAC AGCTACCACA GTCCTCCTGG TATTATTTCT CTGAGGCTGC CCACTCCCCA

640 6 CAACAG

TI GII ACA GGI ICI GGI CAI GCA AGC ICI ACC CCA GGI GGA GAA AAG GAG 641 EXON 2

6/6 ACT TOG GCT ACC CAG AGA AGT TOA GTG CCC AGC TCT ACT GAG AAG AAT GCT GTG AGT ATG ACC AGC AGG

GTA CTC TCC AGC CAC AGC CCC GGT TCA GGC TCC CCA CCA CTC AGG GAC AGG ATG TCA CTC TGG CCC CGG

TCC CAG TCA CCA GGA CCA CGG AAC CAG CTI CAG GTI CAG CTG CCA CCT GGG GAC AGG ATGTCA CCT CGG

968 GCC CTG GGC TCC ACC ACC GCG CCA GCC CAC GAT GTC ACC TCA GCC CCG GAC AAC AAG CCA GCC CCG GG

-- → TANDEM REPEAT

Fig. 3 Cont 'd(2)

INTERNATIONAL SEARCH REPORT

International Application No PCT/GB 90/02020

I. CLASS	FICATION OF SUBJECT MATTER (if several classif	ication symbols apply, indicate all) ⁶	
According	- International Patent Classification (IPC) or to both Natio	onal Classification and IPC	12 7 21 /00
IPC ⁵ :	C 07 H 21/04, C 07 K 13/0	0, C 12 N 15/12, C	12 P 21/00,
IPC :	C 12 Q 1/68, G 01 N 33/57	4, C 07 K 15/00	
II. FIELDS	SEARCHED		
	Minimum Documen		
Classificatio	n System	Classification Symbols	
5		•	
IPC ⁵	C 07 K, C 12 N, C 1	2 P, C 12 Q	
	Documentation Searched other to	han Minimum Documentation	
	to the Extent that such Documents	are included in the Fields Searched	
III. DOCU	MENTS CONSIDERED TO BE RELEVANT		I D. I
Category *	Citation of Document, 11 with indication, where app	ropriate, of the relevant passages 12	Relevant to Claim No. 13
х	WO, A, 8805054 (IMPERIAL	CANCER RESEARCH	6
	TECHNOLOGY)		
ļ.	14 July 1988 see the whole documen	t: especially	
	claims	c, capcording	
1	cited in the application		
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		mistry vol 265	1,2,6-9
P,X	Journal of Biological Che no. 10, 5 April 1990,	The American	1,2,0
	Society for Biochemis	try and Molecular	
	Biology, Inc., (US),	cri una nerecara	·
	M.J.L. Ligtenberg et	al.: "Episialin,	
	a carcinoma-associate	ed mucin, is	
·	generated by a polymo	orphic gene	
	encoding splice varia	ints with	
	alternative amino ter	mini", pages 5573-	
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* Specia	categories of cited documents: 10	"T" later document published after to priority date and not in confl	
WAT don	ument defining the general state of the art which is not sidered to be of particular relevance	cited to understand the princip	e or theory underlying the
"E" ear	ier document but published on or after the international	avu deciment of particular rainvan	ce; the claimed invention
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late	r than the priority date claimed	"4" document member of the same	patent family
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	EUROPEAN PATENT OFFICE	MEST.	TA ZELAAK

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P,X P,X	Journal of Biological Chemistry, vol. 265, no. 25, 5 September 1990, The American Society for Biochemistry and Molecular Biology, Inc., (US), S.J. Gendler et al.: "Molecular cloning and expression of human tumorassociated polymorphic epithelial mucin", pages 15286-15293 see figure 1 Journal of Biological Chemistry, vol. 265, no. 25, 5 September 1990, The American Society for Biochemistry and Molecular Biology, Inc., (US), M.S. Lan et al.: "Cloning and sequencing of a human pancreatic tumor mucin cDNA", pages 15294-15299 see the whole article	1,2,6-9
P,X	no. 25, 5 September 1990, The American Society for Biochemistry and Molecular Biology, Inc., (US), S.J. Gendler et al.: "Molecular cloning and expression of human tumorassociated polymorphic epithelial mucin", pages 15286-15293 see figure 1 Journal of Biological Chemistry, vol. 265, no. 25, 5 September 1990, The American Society for Biochemistry and Molecular Biology, Inc., (US), M.S. Lan et al.: "Cloning and sequencing of a human pancreatic tumor mucin cDNA", pages 15294-15299	1,2,6-9
	no. 25, 5 September 1990, The American Society for Biochemistry and Molecular Biology, Inc., (US), M.S. Lan et al.: "Cloning and sequencing of a human pancreatic tumor mucin cDNA", pages 15294-15299	
P.A		ļ
	WO, A, 9005142 (IMPERIAL CANCER RESEARCH TECHNOLOGY) 17 May 1990 see abstract and claims cited in the application	1-10,14

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET
X
V. TOBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE !
This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons: 1. Claim numbers XX, because they relate to subject matter not required to be searched by this Authority, namely: Claims 11-13
Pls. see Rule 39.1 (iv) - PCT:
Methods for treatment of the human or animal body by surgery or therapy, as well as diagnostic methods.
2. Claim numbers, because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claim numbers because they are dependent claims and are not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).
VI. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING ²
This International Searching Authority found multiple inventions in this international application as follows:
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.
2. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:
No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:
4. As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.
Remark on Protest The additional search fees were accompanied by applicant's protest.
No protest accompanied the payment of additional search fees.

PROPERTY.

35.

ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.

GB 9002020 SA 43255

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 07/05/91

The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Patent document Publication ed in search report date		Publication date
₩0-A- 8805054	14-07-88	AU-A- 110398 EP-A- 034125 JP-T- 250182	2 15-11-89
WO-A- 9005142	17-05-90	None	
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For more details about this annex : see Official Journal of the European Patent Office, No. 12/82